

Volume 15 – December 19, 2021 A Newsletter for Pyraloidea Fans

Inside:

GlobIZ News NZ:On the brink of extinction European Alp odontiines News From... In Search of... About Pyraloidologists... From the Web.... Publications....

Editorial

This issue of the newsletter includes contributions by pyraloid enthusiasts worldwide on a broad diversity of topics, including conservation, climate change, ecology, phylogenetics, new distributions, and taxonomic identities, among others. There are two contributions that speak to the potential of climate change to affect pyraloid species: species in New Zealand that are on the precipice of extinction, and observations on a small, darkly colored, high elevation odontiine genus in the European Alps. Two phylogenies published this year are highlighted: *Ostrinia*, an economically important genus with some species that are serious pests of food crops worldwide, and a higher-level phylogeny of Pyraloidea using mitogenomes. This issue also includes an introduction to Hector Vargas a Chilean who has published on pyraloids and their biology from the southern Western Hemisphere. A new book titled "A global comprehensive check-list of the Phycitinae," a contribution of about 500 pages by Guillaume H. C. Leraut, is highlighted. New in this issue is a "In Search Of" section, for lepidopterists seeking pyraloid specimens or images.

Congratulations to two pyraloidologists who successfully completed their PhDs: Dr. **Qi Muji**e, Nankai University, Tianjin, China, and Dr. **Jae-Ho Ko**, Incheon National University, Incheon, Korea! Also, congratulations to **Joël Minet**, who is now retired and continues as Emeritus Professor at the Muséum National d'Histoire Naturelle, Paris, France.

Matthias Nuss provided an update to Globiz as always. Corrections, inclusions, or questions are welcomed; do not hesitate to contact any one of the authors on the website (www.pyraloidea.org). I was contacted several times this year about entries in Globiz. The pyraloid community owes a great debt of gratitude to **Richard Mally**, who continues to generously update the pyraloid database.

This year was an unusual year for me. Besides being on Zoom (or other communication platforms) frequently like everyone else, I made several presentations as pre-recorded talks. I finished my two years as President of the Lepidopterists' Society and gave a Presidential Address where I spoke about my mentors and mentoring younger people (especially about interns and the NMNH pyraloid collection) and gave the keynote speech at the Latino/Hispanic Symposium at the Entomological Society of America about my career, which also included a section about how I got started in moths [some previously appeared in Solis, 2021: President's Letter. Newsletter of the Lepidopterists' Society 62 (4): 168-172] that will be on YouTube eventually.

Finally, in November I was incorporating material into the pyraloid collection when a group came by with the NMNH Entomology Collection Manager, Floyd Shockley. They turned out to be journalists and photographers from National Geographic on-line. They took the only picture I have of me working in the collection after almost 40 years, and we spoke about chrysaugines, particularly the sloth moths, because this was where I happened to be working when they came by (*A rare look inside the Smithsonian's secret storerooms* by Bill Newcott, photos by Rebecca Hale, available on-line at their site).

This was another difficult year for many due to the pandemic, so thank you to those who brought research papers to my attention throughout the year, and everyone who was able to send in items and/or images for the newsletter.

M. Alma Solis

GlobIZ News 2021

The Global Information System on Pyraloidea (GlobIZ) underwent some revisional checks this year. Since the last newsletter, the number of valid species included in the database increased by 162 (+ 55 synonyms). Altogether, there are 26,748 pyraloid names for 2,118 genera (+ 1,438 synonyms) and 16,621 species (+ 6,571 synonyms). Numerous nomenclatural changes account for the numerical changes. The updated table below provides an overview by subfamily. I very much like to thank all who contributed editing data to GlobIZ, especially Richard Mally who again spent much time in 2021 for carefully checking possible gaps in the database.

Matthias Nuss

	genera		species	
	valid synonyms		valid synonyms	
Chrysauginae	130	61	399	129
Epipaschiinae	95	68	727	172
Galleriinae	63	62	260	117
Phycitinae	676	401	3,501	1,560
Pyralinae	136	109	1,298	399
Acentropinae	71	38	797	214
Crambinae	177	124	2,083	1,098
Erupinae	3	4	38	5
Glaphyriinae	75	56	509	197
Heliothelinae	3	3	29	14
Hoploscopinae	2	4	46	2
Lathrotelinae	5	7	42	9
Linostinae	1	0	4	2
Midilinae	10	4	57	12
Musotiminae	23	8	208	26
Odontiinae	87	39	386	143
Pyraustinae	172	106	1,284	633
Schoenobiinae	29	17	240	99
Scopariinae	20	24	587	208
Spilomelinae	340	303	4,126	1,532
	2 1 1 0	1 / 20	16 621	6 571
	2,118	1,438	16,621	. 0,3/1

Some New Zealand Pyraloidea on the brink of extinction

Robert J. B. Hoare Landcare Research, Auckland, New Zealand

For its size, New Zealand has a diverse and unusual fauna of Pyraloidea, with 250 currently recognised species in 11 subfamilies (one species is unplaced to subfamily), of which a remarkable 234 species are endemic (93.6%). The fauna is overwhelmingly dominated by Crambinae (81 species) and Scopariinae (134 species), with the majority of species occurring in the South Island, where major radiations have taken place in the alpine zone.

New Zealand's landscapes and ecology have changed drastically since humans arrived and especially since European settlement over the last 200 years. By and large, the country's insect fauna has fared somewhat better than its devastated avifauna, and no species of Lepidoptera have been declared extinct. However, one formerly locally common geometrid moth, Xanthorhoe bulbulata (Guenée, 1868), has not been seen since 1991, despite intensive searching, and seems very unlikely now to be rediscovered. It has recently become clear that other species occurring in similar habitats to X. bulbulata are severely threatened, and several pyraloid moths are perhaps the most endangered.

The habitats of concern are short-turf areas rich in native herbs occurring in a wide range of settings from coastal dunes to intermontane basins and low alpine areas of the South Island, especially to the east and south of the Southern Alps. The turfs may be dry or seasonally wet, and often occur in mosaics of shrubland and native tussock grassland;

lakesides and river flats are classic examples at the moister end of the scale, saltpans at the drier end. These habitats have become increasingly overwhelmed by exotic grasses and weeds, accidentally or deliberately introduced over many decades of land-use change relating to settlement and agriculture. Rabbits and hares (also introduced) can be abundant and damage these open areas extensively, but ironically, may in some places help to keep down exotic grasses and allow native herbs to thrive. It is possible that moderate sheep-grazing may also be beneficial in some situations, but more research is needed. Climate change has warmed New Zealand by about 1°C over the last 100 years and is a likely exacerbating factor in the ongoing degradation of these delicate turfs. The interactions are complex and are not adequately understood from the perspective of Lepidoptera conservation.

With Brian Patrick, I have been attempting to assess the status of four endemic pyraloid moths confined to these short-turf habitats; all were formerly widespread and locally common in the eastern and southern South Island. These are: Maoricrambus oncobolus (Meyrick, 1885) (Crambidae: Crambinae), Pyrausta comastis (Meyrick, 1884) (Crambidae: Pyraustinae), Delogenes limodoxa Meyrick, 1918 (Pyralidae: Phycitinae) and Sporophyla oenospora (Meyrick, 1897) (Phycitinae) (Figs 1-4). Apart from P. comastis, all are currently placed in monotypic endemic genera, though Maoricrambus is closely related to Orocrambus and may eventually prove to be a synonym. These species have never been reared and the life histories are unknown, so a critical component of the research was to rediscover populations in the hope that larvae could be found later, and their ecology elucidated. Maoricrambus, in common with its rel-



Figs 1-4. Endangered New Zealand pyraloids: 1. *Maoricrambus oncobolus*; 2. *Pyrausta comastis*; 3. *Delogenes limodoxa*; 4. Sporophyla *oenospora*.

atives, is almost certainly a monocot-feeder, whilst the other three are strongly suspected to be associated with native herbs. All four are day-flying and were formerly rather easily observed in suitable habitat at the right season. None of the four species has ever been reported on the popular *iNaturalist* NZ recording platform.

The results of our surveys in many former sites for these species over the last two summers (2019-2021) have been deeply worrying (Fig. 5). We found no specimens at all of three of the species; only a single population of Delogenes limodoxa was found in a complex of dwarf shrubland and herbfield on quartz-rich dunes on the south coast near Invercargill, where 7 specimens were observed by Brian Patrick. The herb layer here is relatively diverse, so finding larvae and confirming the hostplant(s) will be a challenge. The species has also been found in the last 5 years by Roland and Renate Wöger further east near Waikawa, and by Brian Lyford in the Otago Lakes area, so is likely hanging on very locally in a number of places.

The status of the other three species appears critical. Most of the sites we visited were in protected areas, but nonetheless quite substantial changes have occurred over the last 10 to 15 years; many sites have a notably longer sward and have been much more extensively invaded by weeds and exotic grasses than before. Sporophyla oenospora was formerly found coastally as well as in dry inland sites, including saltpans in Central Otago, but it has not been seen anywhere since 2008. Pyrausta comastis, an inhabitant of wetter swards or short grass, was still relatively common in 2010 at a single site in Fiordland but has not been seen since that date. Maoricrambus oncobolus occurred very locally in estuarine situations, as well as on intermontane river flats, and was still common at one remote inland site in 2007; it is possible that recent surveys at known sites were too late in the season, so there is hope that this moth may still survive.



Fig. 5. Red Tarns, 1200m a.s.l., near Mt Cook Village, South Island, New Zealand (Jan 2020). *Delogenes limodoxa* and *Pyrausta comastis* were found commonly here on lakeside swards by Alfred Philpott in 1929. The habitat looks superficially promising, but weeds are present and these pyraloids seem to have disappeared.

We concluded that *S. oenospora* and *P. comastis* are on the brink of extinction, and chances to save them are rapidly diminishing, if they still exist. The decline of these moths was barely noticed until a couple of years ago when this project was initiated, due to a severe lack of recording in critical areas of the South Island and a general lack of specialist lepidopterists with the experience to search for and recognise the moths.

On a more positive note, these four rare species should be considered extremely sensitive indicators of habitat change, and their rapid decline has alerted us to a need for urgent research into the ecology and management of New Zealand's threatened short-turf habitats. Other specialist moths are still locally common in these habitats; they are perhaps more adaptable, but they should not be assumed to be safe. We sincerely hope there is time to prevent these species suffering a similar fate.

This research was funded by the New Zealand Department of Conservation, and we are very grateful to Eric Edwards, Tara Murray and Samantha Gale for their help and support.

Metaxmeste (Odontiinae) in the European Alps

Matthias Nuss Senckenberg Museum für Tierkunde, Dresden, Germany

This summer, I was hiking in the Ötztal Alps, an area famous for Ötzi, the Iceman who was killed with an arrow 5,300 years ago. Brilliant weather conditions allowed spectacular views on the glacier scenery, but note that the ice shields are rapidly shrinking, releasing bare, smoothly polished rock (Fig. 1). The alpine meadows seen in the foreground are home to two alpine odontiines, *Metaxmeste* phyrgialis and M. schrankiana (Figs. 2-3). Adults of both species fly during the day, close to the ground, and are usually seen sunbasking on bare ground. Schmid (2019) calls M. phyrgialis (Fig. 2) probably the most common alpine micro-moth species in the Alps and mentions that its larvae are polyphagous. Less common is M. schrankiana (Fig. 3) whose larvae are known to feed on Vaccinium (Ericaceae), Geum, and Dryas (Rosaceae).

Returning home, I checked published

records, for example Sinev (2019), and became fascinated with the distribution patterns of the two species, which are scattered over several boreal and mountainous regions in Eurasia and are isolated by widely extended lowlands. *M. phyrgialis* even has a counterpart in Colorado, *M. nubicola* (See MPG web site). The wing pattern of these two species looks much the same to me.



Fig. 1. Scenery of the Gaisberg (left) and Rotmoos glaciers as seen from the Mt. Hohe Mut (2,653 m).



Fig. 2. *Metaxmeste phyrgialis* in the Gaisbergtal at about 2,400 m.

Thanks to correspondence with Alma Solis, Zhaofu Yang, Jean-François Landry and Paul Opler, I was able to quickly analyse *nubicola*'s DNA-barcode against Eurasian specimens, which resulted in a sequence distance well over 2%. Whether *M. phrygialis* remains the most common alpine micro-moth over the next decades will greatly depend on how climate change proceeds. The hibernating habit of this species depends on snow cover, but the snow line will move upwards and there will be more rain during the winter.

References

Schmid, J. 2019: Kleinschmetterlinge der Alpen. Verbreitung, Lebensraum, Biologie. Haupt Verlag, Bern. 800 S.

Sinev, S. Ju. 2019: Catalogue of the Lepidoptera of Russia. Zoological Institute, St. Petersburg. 448 S.



Fig. 3. *Metaxmeste schrankiana* below summit of Mt. Hohe Mut at about 2,600 m.

NEWS FROM.....

Zhaofu Yang New Ostrinia Phylogeny

Yang, Z., D. Plotkin, J.-F. Landry, C. Storer, & A. Kawahara. 2021. Revisiting the evolution of *Ostrinia* moths with phylogenomics (Pyraloidea: Crambidae: Pyraustinae). Systematic Entomology. 46: 827-838.

The crambid moth genus *Ostrinia* contains multiple agricultural pests, and its classification. We inferred a molecular phylogeny of *Ostrinia* using a phylogenomic dataset containing 498 loci and 115, 197 nucleotide sites and examined whether traditional morphological characters corroborate our molecular results.

Our results strongly support the monophyly of one of the Ostrinia species groups (Fig. 1 from research paper on next page), but surprisingly do not support the monophyly of the other two. Based on the extensive morphological examination and broadly representative taxon sampling of the phylogenomic analyses, we propose a revised classification of the genus, defined by three species groups (Ostrinia nubilalis species group, Ostrinia obumbratalis species group, and Ostrinia penitalis species group), which differs from the traditional classification of Mutuura & Munroe (1970). Morphological and molecular evidence reveal the presence of a new North American species, Ostrinia multispinosa Yang sp.n., closely related to O. obumbratalis. Our analyses indicate that the Ostrinia ancestral larval host preference was for dicots, and that O. nubilalis (European corn borer) and Ostrinia furnacalis (Asian corn borer) independently evolved a preference for feeding on monocots (i.e., maize). Males of a few Ostrinia species have enlarged, grooved midtibiae with brush organs that are known to attract females to increase mating success during courtship, which may represent a derived condition. [abstract slightly modified].

Mujie Qi and Houhun Li Pyraloidea Mitogenomes and phylogeny

Liu, Xiaomeng, M.Qi, H. Xu, Z.Wu, L. Hu, M. Yang, & H. Li. 2021. Nine mitochondrial genomes of the Pyraloidea and their phylogenetic implications (Lepidoptera: Pyraloidea). Insects. 12: 1039. https://doi.org/10.3390/ insects12111039

We sequenced and annotated nine complete mitogenomes for Pyraloidea, and further performed various phylogenetic analyses, to improve our understanding of mitogenomic evolution and phylogeny of this superfamily (Fig. 3 from research paper on next page). TGene content and arrangement were highly conserved and are typical of Lepidoptera. Based on the hitherto most extensive mitogenomic sampling, our various resulting trees showed generally congruent topologies among pyraloid subfamilies, which are almost in accordance with previous multilocus studies, indicating the suitability of mitogenomes in inferring high-level relationships of Pyraloidea. However, nodes linking subfamilies in the "non-PS clade" were not completely resolved in terms of unstable topologies or low supports, and future investigations are needed with increased taxon sampling and molecular data.

Unexpectedly, *Orybina* Snellen, represented in a molecular phylogenetic investigation for the first time, was robustly placed as basal to the remaining Pyralidae taxa across our analyses, rather than nested in Pyralinae of Pyralidae as morphologically defined. This novel finding highlights the need to reevaluate *Orybina* monophyly and its phylogenetic position by incorporating additional molecular and morphological evidence. [abstract slightly modified].



Fig. 1: Maximum likelihood tree of *Ostrinia* estimated in iq-tree using the anchored hybrid enrichment dataset. Support values are ufboot/sh-alrt.. Nodes of importance are numbered in bold. The ancestral state reconstructions (ASRs) of larval host plant associations are represented as pies on ingroup nodes. The ASRs of the male midtibiae are indicated with capital letters following tip labels as follows: (L) enlarged, (S) small.



Fig. 3: Resulting trees constructed with MrBayes for five datasets. (A) The whole BI tree of PCG123 dataset, and position of the *Orybina* Snellen is emphasized with red clade and polygon; (B–E) highlight the partial BI trees ("*non-clade*") constructed using the datasets of PCG12, PCG12R, PCG123R and PCGAA, respectively.

Stephen Sutton Pyraloid Moths of Borneo (PoB), Volume 2

Everything is ready to submit to the publisher for printing, except that the key process of loading text and images to the website and generating the QR codes to link the printed volume to the site is on hold for the moment. We are searching for a new software programmer. As of 17 March 2021, there were 798 species entries including all records for all subfamilies of the Crambidae in Borneo (or highly likely to occur) and genera of the Spilomelinae in A-Z to *Gadessa*. The rest of the Spilomelinae will comprise Vol 3.

Jean-Michel Maes

Pyraloidea in iNaturalist

Perhaps a way to increase knowledge about pyraloids is cooperating with iNaturalist, a platform where amateurs can upload observations of plants and animals. So far there are 413,539 pyraloid observations, 99 % with pictures [numbers from December 2020], representing 3,518 species. In the United States: 238,159 observations, 1,051 species; South America: 19,601 observations, 819 species; Mexico: 12,524 observations, 470 species; Nicaragua: 192 observations, 79 species; and Costa Rica: 3,049 observations, 277 species. Central and South America are not well represented, but the potential is huge. Few people take pictures of pyraloids because very few can identify small moths, even if many are beautiful species. iNaturalist could be used as a taxonomic platform, and perhaps if pyraloid specialists would identify species where possible, it would boost enthusiasm about pyraloids among amateurs.

Clifford Ferris First records of *Maruca vitrata* (Fabricius, 1787) (bean pod borer) in Arizona

Two specimens of this agriculturally important pest species were collected in UV moth traps on successive nights (Fig. 1). The first from a trap placed along Harshaw Creek, SE of Patagonia, Sta. Cruz Co., Arizona [N31°30.893', W110°42.241'] 4270' (1302m)], 7 September 2021. The second specimen was taken in Gardner Canyon, Pima Co., Arizona [N31°43.627', W110°41.833'] 4805' (1465m), 8 September 2021. This moth has been reported once from Louisiana and several times from peninsular Florida, but not previously from the southwestern United States. The two specimens are presumed to be migrants from Mexico since there is no commercial bean cultivation in the two areas where the moths were collected. A strong diurnal migration of snout butterflies and yellow pierids was occurring on the two dates noted above.



Fig. 1. Maruca vitrata collected in Arizona

Bob Heckford Biology of *Pyrausta*

Heckford, R. J. & S. D. Beavan. 2021. On the biology of *Pyrausta purpuralis* (Linnaeus, 1758) and its comparison with *Pyrausta ostrinalis* (Hübner, 1793) and *Pyrausta aurata* (Scopoli, 1763) (Lepidoptera: Pyralidae). Entomologists' Gazette. 72: 85-118.



Pyrausta aurata ©S. D. Beavan & R. J. Heckford



Pyrausta ostrinalis ©S. D. Beavan & R. J. Heckford



Pyrausta purpuralis©S. D. Beavan & R. J. Heckford

Scott Miller Ecological studies including pyraloids

Basset, Y., L. R. Jorge, P. T. Butterill, G. P. A. Lamarre, C. Dahl, R. Ctvrtecka, S. Gripenberg, O. T. Lewis, H. Barrios, J. W. Brown, S. Bunyavejchewin, B. A. Butcher, A. I. Cognato, S. J. Davies, O. Kaman, P. Klimes, M. Knížek, S. E. Miller, G. E. Morse, V. Novotny, N. Pongpattananurak, P. Pramual, D. L. J. Quicke, W. Sakchoowong, R. Umari, E. J. Vesterinen, G. D. Weiblen, S. J. Wright, and S. T. Segar. 2021. Host specificity and interaction networks of insects feeding on seeds and fruits in tropical rainforests. Oikos. 130(9):1462-1476.

This meta-analysis includes Pyralidae, but not Crambidae because they were not common enough in fruits and seeds to make the threshold for analysis. [modified abstract following]. Host specificity was greater among seed eaters than pulp-eaters and for insects feeding on dry fruits as opposed to insects feeding on fleshy fruits. Plant species richness within plant families did not influence insect host specificity. Some subnetworks were extremely specialized, such as those including Tortricidae and Bruchinae in Panama. Plant phylogenetic distance, plant basal area and plant traits (fruit length, number of seeds per fruit) had important effects on several network statistics in regressions weighted by sampling effort. Our study emphasizes the duality between seed dispersal and seed predation networks in the tropics, as key plant species differ, and host specificity tends to be low in the former and higher in the latter. This underlines the need to study both types of networks for sound practices of forest regeneration and conservation.

Volf, M., J. E. Laitila, J. Kim, L. Sam, K. Sam, B. Isua, M. Sisol, C. W. Wardhaugh, F. Vejmelka, S. E. Miller, G. D. Weiblen, J.-P. Salminen, V. Novotny, and S. T. Segar. 2020. Compound specific trends of chemical defences in *Ficus* along an elevational gradient reflect a complex selective landscape. Journal of Chemical Ecology. 46(4):442-452.

This study included species of Glyphodes, Talanga, and a scopariine. [modified abstract follows] Elevational gradients affect the production of plant secondary metabolites through changes in both biotic and abiotic conditions. We analyzed the correlation of alkaloids and polyphenols with elevation in a community of nine Ficus species along a continuously forested elevational gradient in Papua New Guinea. Insect community structure was affected mainly by alkaloid concentration and diversity. Although our results show an elevational increase in several groups of metabolites, the drivers behind these trends likely differ. Flavonoids may provide figs with protection against abiotic stressors. In contrast, alkaloids affect insect herbivores and may provide protection against mammalian herbivores and pathogens.

Seifert, C. L., L. R. Jorge, M. Volf, D. L. Wagner, G. P. A. Lamarre, S. E. Miller, E. Gonzalez-Akre, K. J. Anderson-Teixeira, and V. Novotný. 2021. Seasonality affects specialization of a temperate forest herbivore community. Oikos. 130(9):1450-1461.

This study did not include any Pyraloidea species, but it shows what you can do with larvae if you have a good DNA barcode library.

Alma Solis Various pyraloids

Diatraea postlineella was previously only known from the type described by Schaus from Guatemala. This species came to my attention when I taught a two-week course on various aspects of Pyraloidea in El Salvador in 2012. With additional material many years later, I was able to study morphological variation, including the female for the first time (Solis et al., 2021). My collaborators provided the new host plant data, and a COI barcode identity for the first time. This species had been misidentified as *D. grandiosella* by Guatemalan researchers.

Jenny Phillips and I came upon *Crypto-cosma*, a group with morphologically interesting adult moths (i.e., reduced mouthparts and a cataclystiform wing pattern), in the USNM collection while working on the Acentropinae of the Guanacaste Conservation Area (Costa Rica). We synonymized *Guyanymphula* with *Cryptocosma*, and transferred the genus from Acentropinae to Glaphyrinae (Solis & Phillips, 2021).

Unusually, and because of the pandemic, I collaborated with an intern, Jacob Bethin, via Zoom. He discovered aquatic Lepidoptera in a collection of the River Weed plant family at his university. He photographed the specimens, and we discussed aquatic acentropine larvae and pupae (see pupal image to the right) every Friday this summer! I encouraged him to write a paper on the unusual morphological structures (Bethin et al., 2021). He has applied for an NSF grant to work on these amazing pyraloids.

I was involved in a multi-authored paper, a "minimalist revision," about braconids that are parasitoids of Costa Rican pyraloid larvae (Sharkey et al., 2021). This paper caused quite a stir because the first author included many new species diagnosed and based only on COI bins. I made/confirmed identifications, including of various invasive species in the United States. *Cydalima perspectalis*, the box tree moth, was finally found in the United States, and there were new county records in Texas of the cactus moth, *Cactoblastis cactorum*, as it spreads westward.

References

Bethin, J., M. A. Solis, & R. K. Krell. 2021. The Undiscovered Frontier of Aquatic Moth Caterpillars on River Weeds. Newsletter of the Lepidopterists' Society. 63 (3): 143-146.

Sharkey, M. J., D.H. Janzen, W. Hallwachs, E. Chapman, M.
A. Smith, T. Dapkey, A. Brown, S. Ratnasingham, N. Suresh,
M. Ramya, K. Perez, M. Milton, P.D.N. Hebert, S. R. Shaw, R.
N. Kittel, M. A. Solis, M. A. Metz, P. Goldstein, J. W. Brown,
D.L.J. Quicke, C. Van Achterberg, B.V. Brown, & J. Burns.
2021. Minimalist revision and description of 411 new
species in 11 subfamilies of Costa Rican braconid parasitic
wasps, including host records. Zookeys. 1013:1-165.doi.
org/10.3897/zookeys.1013.55600

Solis, M. A., & E. Phillips. 2021. Hidden Jewel: *Cryptocosma* Lederer (Lepidoptera: Pyraloidea: Crambidae), its transfer to the Glaphyriinae, and the synonymy of *Guyanymphula* Heppner. Proceedings of the Entomological Society of Washington. 123(1): 217-229. doi.org/10.4289/0013-8797.123.1.217

Solis, M. A., S. J. Scheffer, M. Lewis, & P. Rendon. 2021. *Dia-traea postlineella* Schaus (Lepidoptera: Crambidae) from Guatemala: molecular identity and host plant. Proceedings of the Entomological Society of Washington. 123(3): 638-651.



IN SEARCH OF....

*Syllepte incomptalis Hübner from Surinam Richard Mally, Matthias Nuss, & Alma Solis

Syllepte is probably the most troublesome dustbin genus in Spilomelinae, with 204 species worldwide largely varying in forewing length, wing pattern elements, and genitalia characters. Munroe (1995) indicated that almost all New World species were misplaced, with the exception of the type species, S. incomptalis Hübner, 1819–1823, described from Surinam, and *S. nitidalis* (Dognin, 1905) described from Ecuador (Figs. 1-2). Therefore, it is labor-intensive to properly place a single species, and even more challenging to understand what a true Syllepte looks like, since the original specimens of Hübner's descriptions were lost in a fire in Vienna during the 1848 revolution. In the original description, the species is figured in colour (pl. 50, figs. 285-286) (Fig. 1) and the written description, despite mentioning similar taxa, is limited to the wing pattern which is straw yellow with a reddish hue. Looking at these few facts, the identity of S. incomptalis remains dubious.

When looking carefully at Hübner's plates, it becomes evident that the specimens he illustrated are in correct proportion to each other, which helps to estimate the size of *S. incomptalis*. In Hübner (1819–1823) Plate 50, next to *S. incomptalis*, is an illustration of *Melanis lycea* (figs. 283–284), which has a forewing length of about 22 mm (compare at https://www.butterfliesofamerica.com/L/melanis_lycea_types.htm). With *S. incomptalis* being less than half the size of *M. lycea*, its forewing length can be estimated to be 10 mm. This is also consistent with the comparable size of *Anomis erosa* (pl. 50, figs. 287–



Fig. 1. Hübner's (1819-1823) plate 50 with the two illustrations of a male *Syllepte incomptalis* (see also Pyraloid Planet, v.13, p. 16)



Fig. 2. S. nitidalis (Dognin) syntype, USNM.



Fig. 3. *S. nitidalis* (Dognin), syntype, posterior abdominal segments, USNM.



Fig. 4. *S. nitidalis* (Dognin) male genitalia, syntype, USNM.

288), with a forewing length of 12–13 mm (Powell & Opler 2009). The image and supposed size of the wing length are very similar to *S. nitidalis* (See adult and genitalia syntype images, Figs. 2-4) with the strong possibility that *S. incomptalis* is the same or closely related to *S. nitidalis* (Figs. 3-4).

The USNM has material of *S. nitidalis* from Mexico, Guatemala, Costa Rica, Ecuador, Peru, Venezuela, and Cuba, and several undescribed, closely related species from Ecuador, Bolivia, Peru, Costa Rica, and Dominica. We would like to ask any pyraloid enthusiast to carefully check her/his Neotropical material for possible *Syllepte incomptalis* or *nitidalis* from Surinam or any other country in the Guiana Shield of South America. We would be very happy to borrow those specimens for study.

References

Hübner, J. 1819–1823 [imprint "1823"]: Zuträge zur Sammlung exotischer Schmettlinge [sic], bestehend in Bekundigung einzelner Fliegmuster neuer oder rarer nichteuropäischer Gattungen. Augsburg. [1]–[3]–4-6– [7]–8–32–[33]–[40], pls [36]–[69]. Plate 50: https://www. biodiversitylibrary.org/item/45768#page/103/mode/1up

Munroe, E. 1995. Pyraustinae, pp. 53-79 In: J.B. Heppner, ed. Check List Part 2: Atlas of Neotropical Lepidoptera, Brill/Flora and Fauna Books, Gainesville, FL. 243 pp.

Powell, J. A., & Opler, P. A. 2009: Moths of Western North America. University of California Press. https://doi. org/10.1525/9780520943773

*Pyralinae and Australian Chrysauginae immature stages James Hayden

I started to score a morphological matrix of Pyralidae earlier this year. It started from a dichotomous key to pyraloid familylevel taxa that I have been working on. The key emphasizes external characters to aid sorting specimens in the collection. I encountered problems with some genera with unusual combinations of characters, so I realized that scoring a whole matrix for Pyralidae was necessary, incorporating as much morphological evidence as possible. I started it during the pandemic lockdown when my resources were limited to the collection where I work. I showed results of the analysis at The Lepidopterists' Society meeting on August 20th. The results are very preliminary, but interesting.

Some of the problematic genera still do not fit in any of the five current subfamilies. Pyralinae especially is poorly defined, which is not news (Solis and Mitter 1992, Solis and Shaffer 1999). Even if more complete results is the same as the status quo, it would be useful to publish a matrix-based key to pyralid taxa for the benefit of non-specialists.

I would like to examine more immature stages, so I am interested in borrowing or exchanging larvae or pupae. Of the following, the larval hosts of some have been published, or perhaps the larvae have been collected but not reported, as the moths are not uncommon: *Sindris, Episindris* and related genera (African), *Sacada, Datanoides* (African and Asian), *Propachys, Orybina,* or *Trebania* (Asian), *Centropseustis astrapora* (see Meyrick 1890), and any of the Australian "chrysaugines" (*Anemosa, Hednotodes, Polyterpnes*).

Please write to me if you would like to collaborate at Florida Department of Agriculture and Consumer Services, Division of Plant Industry, 1911 SW 34th Street, Gainesville, Florida, 32608, USA, or email at: james. hayden@fdacs.gov.

References

Meyrick, E. 1890. Descriptions of additional Australian Pyralidina. Proceedings of the Linnean Society of New South Wales (ser. 2) 4: 1105–1116.

Solis, M. A. and C. Mitter. 1992. Review and preliminary phylogenetic analysis of the subfamilies of the Pyralidae (sensu stricto) (Lepidoptera: Pyraloidea). Systematic Entomology 17: 79–90.

Solis, M. A. and M. Shaffer. 1999. Contribution towards the study of the Pyralinae (Pyralidae): historical review, morphology, and nomenclature. Journal of the Lepidopterists' Society 53(1): 1–10.



Orybina flavipaga Walker, Taiwan

*Image of live sloth moth Akito Kawahara

An image of a live sloth moth is needed for a Lepidoptera book I am working on wit Jim Miller.

About Pyraloidologists... Dr. Héctor Vargas

I am an Associate Professor at Universidad de Tarapacá, Arica, in the northernmost part of Chile (Fig. 1).



Fig. 1

I am very interested in the systematics and natural history of local Lepidoptera associated with native plants of the extremely arid environments of the Atacama Desert and Andes of northern Chile (See Vargas, p. 17 & p. 19 in Publications below). Although this geographic area harbors low plant diversity, some representatives of Lepidoptera occur here, among them a few members of Pyraloidea. For instance, just a few minutes ago, I wrote labels for and pinned a gray Phycitinae moth (Fig. 2), which I collected last month as a larva on inflorescences of the wonderful shrub Miconia acuminata (Asteraceae) (Fig. 3) near the small village of Socoroma at about 3400 m elevation in the Andes.



Fig. 2





Because many families of moths in northern Chile have been poorly studied, the only certainty I have for now, regarding the above-mentioned specimen, is that it will represent a very interesting puzzle before reaching an accurate taxonomic identification. I will begin to study it shortly, and soon thereafter will surely contact pyraloid experts to request their always kind help to identify this phycitine.

FROM THE WEB...

iCollections: British and Irish Pyraloidea (Moths) Collection https://data.nhm.ac.uk/ dataset/british-and-irish-pyraloidea-mothscollection

Finally..... New Book

The Revue Française d'Entomologie Générale (Autun, France) in 2021 issued the first systematic and synonymic list at a global level for the Phycitinae: Spécies général des Phycitinae (Lep. Pyraloidea, Pyralidae), A global comprehensive check-list of the Phycitinae. Suppl. to Fasc. 2 (5-6), p. 1-474.

The present work records 5,905 taxa in the tribes Anerastiini, Cryptoblabini, Cabniini and Phycitini with some taxonomic changes. It includes 641 genera, 3452 species, 123 new taxonomic combinations, 3 new genera (Ptocheia, gen. nov., Luqubraphycita, gen. nov., Pylagonia, gen. nov.), as well as 7 new subgenera. Genitalia images of 65 particularly interesting genera are provided, as well as a full bibliography. An executive summary and an abstract are given in English, German, Spanish, Italian, Russian, and Chinese. An update is already planned to include comments received from readers, as well as minor corrections, and a new Russian translation. I hope this book brings attention to the Phycitinae at a global level.

Guillaume H. C. Leraut



Carnasia ulmiarrosorella (Clemens)

A PARAITRE – JANVIER 2021

Spécies général des Phycitinae (Lepidoptera : Pyraloidea, Pyralidae) A global comprehensive checklist of the Pyralidae Phycitinae



A "smattering" of publications

PYRALIDAE

Epipaschiinae

Kim, H., T.-G. Lee, Y.-B. Cha, C.-M. Jang, J.-N. Kim, U. Bayarsaikhan, J.-H. Ko, & Y.-S. Bae. 2021. Review of the genus *Stericta* Lederer (Lepidoptera: Pyralidae: Epipaschiinae) from Korea. Journal of Asia-Pacific Biodiversity. 14: 371-377.

Rong, H., Y. Wang, Q. Mujie, & H. Li. 2021. Taxonomic review of the genus *Lista* Walker, 1859 from China (Lepidoptera, Pyralidae, Epipaschiinae), with descriptions of five new species. Zootaxa. 5081(2): 237-262.

Phycitinae

Akin, K. & E. Seven. 2021. An assessment on genus *Laristania* Amsel, 1951 (Lepidoptera: Pyralidae: Phycitinae) and a new species from Turkey. Zootaxa.4992(2): 169-180.

Alipanah, H. & F. Slamka. 2021. On the genus *Elegia* Ragonot, 1887 (Pyralidae: Phycitinae: Phycitini) from Iran with description of *Elegia kharazii* sp. nov. Zootaxa. 4999(3): 285.

Gumhalter, D. 2021. *Psorosa mediterranella* (Amsel, 1954) (Lepidoptera: Pyralidae, Phycitinae)- a new species for the Croatian pyraloid moth fauna, with an updated checklist. Natura Croatica. 30(1): 37-52. https://doi. org/10.20302/NC.2021.30.4

Nel, J., G. Labonne, & T. Varenne. 2021. *Euzopherodes nigrolineella* Zerny, 1936, bona species, stat. restaur. Description d'*Euzopherodes vapidelloides* sp. n. (Lepidoptera, Pyralidae, Phycitinae). Revue de l'Association Roussillonaise d'Entomologie. 39(1): 29-35.

Pinzari, M. 2020. First description of the male genitalia of *Psorosa lacteomarginata* (A. Costa, 1888) (Lepidoptera: Pyralidae, Phycitinae). SHILAP Revista de Lepidoptera. 48 (189): 109-113.

Pinzari, M. & M. Pinzari. 2020. The genus *Hypsotropa* Zeller, 1848 in Italy and description of *H. aenigmatica* Pinzari & Pinzari, sp. n. (Lepidoptera: Pyralidae, Phycitinae, Peoriini). SHILAP Revista de Lepidoptera. 48 (192): 581-588.

Ren, Y., L. Yang, H. Liu, & H. Li. 2020. Taxonomic review of the genus *Dusungwua* Kemal, Kizildağ, & Koçak, 2020 (Lepidoptera: Pyralidae), with descriptions of six new species and propositions of synonyms. Zootaxa. 4894(3): 341-365.

Suetsugu, K. 2021. No evidence of pollination mutualism between the holoparasitic plant *Mitrastemon yamamotoi* Makino (Mitrastremonaceae) and its herbivore Assara balanophorae Sasaki & Tanaka, 2004 (Lepidoptera: Pyralidae). Pan-Pacific Entomologist. 97(1): 1-5.

Tendolkar, A. A. F. Pomerantz, C. Heryanto, P.D. Shirk, N. H. Patel, & A. Martin. 2021. Ultrabithorax is a micromanager of hindwing identity in butterflies and moths. Frontiers In Ecology and Evolution. 9: 643661. [CRISPR, *Plodia interpunctella*]

Tsvetkov, E. V. 2020. Description of *Pima karatauensis* Tsvetkov, sp. n. from Kazakhstan (Lepidoptera: Pyralidae, Phycitinae). SHILAP Revista de Lepidoptera. 48 (191): 481-485.

Tsvetkov, E. V. 2021. Four new species of the subfamily Phycitinae (Lepidoptera: Pyralidae) from Kazakhstan. Zoosystematica Rossica. 30(1): 116-130.

Vargas, H. 2020. *Ragonotia campodonicoi* Cepeda (Lepidoptera: Pyralidae: Phycitinae) in the Andes highlands of northern Chile, expanded distribution and first documented host plant. Journal of the Lepidopterists' Society. 74(3): 189-192.

Yang, L., H. Liu & Y. Ren. 2021. One new species of the genus *Etielloides* Shibuya, 1928 from China (Lepidoptera, Pyralidae, Phycitinae). Zootaxa. 4990(2): 361-368.

Yepishin, V. 2021. Notes on the taxonomy of Palaearctic *Asclerobia* Roesler, 1969 (Lepidoptera: Pyralidae: Phycitinae) with description of a new species. Zootaxa. 4963(1):149-162.

Pyralinae

Agius, J. & J. Formosa. 2020. Stemmatophora combustalis (Fischer von Röslerstamm, 1842) new to the Maltese Islands (Lepidoptera: Pyralidae). SHILAP Revista de Lepidoptera. 48 (190): 223-224.

Akin, K., C. Gözüaçik, E. Seven, & M. Türkoğlu. 2021. New species and new records of Pyraloid moths (Lepidoptera: Pyraloidea) from Turkey. Zootaxa. 4951(3): 529-540.

Pellinen, M. J., R. Zahiri, & P., Sihvonen. 2020. A new species of *Sacada* Walker, 1862 from Thailand (Lepidoptera: Pyralidae, Pyralinae). Evolutionary Systematics. 4(2): 71-77.

Qi, M. & X. Zuo. 2021. A new species of *Minooa* Yamanaka, 1996 (Lepidoptera, Pyralidae) from Xizang, China. Zootaxa. 4949(1): 191-197.

CRAMBIDAE

Acentropinae

Sexton, C. 2021. Identification and distribution of the *Peterophila fulicalis* species group (Crambidae): taking advantage of citizen science data. Journal of the Lepidopterists' Society. 75(2): 113-127.

Crambinae

Bassi, G. 2021. New genera and species of Afrotropical Ancylolomiini Ragonot, 1889 (Lepidoptera: Pyralidae sensu lato: Crambinae). Revue Suisse de Zoologie. 128(2): 477-486.

Bassi, G., S. Sáfián, G. C. Müller, V. D. Kravchenko, & A. N. Poltavsky. *Ancylogastra*, a new genus of Afrotropical Crambinae, with descriptions of seven new species. Zootaxa. 5052(1): 042-060.

Koren, T. 2021. Further additions to the grass moth (Lepidoptera: Crambidae) fauna of Croatia. Natura Croatica. 30(1): 243-250.

Landry, B. & V. O. Becker. 2021. A taxonomic review of the genus *Diptychophora* Zeller (Lepidoptera, Pyralidae *sensu lato,* Crambinae) in Brazil, with descriptions of three new species. Revue suisse de Zoologie. 128(1): 73-84.

Li, W.C. 2020. A new species of *Crambus* Fabricius, 1798 from China. SHILAP Revista de Lepidoptera. 48 (192): 603-607.

Pyraustinae

Kizildağ, S. 2020. Molecular evaluation of the phylogenetic position of *Loxostege ayhananal* Kemal & Koçak, 2017 from East Turkey (Lepidoptera: Crambidae, Pyraustinae). SHILAP Revista de Lepidoptera. 48 (189): 39-45.

Kizildağ, S. 2021. DNA Barcoding and species delimitation of *Pyrausta* (Lepidoptera: Crambidae, Pyraustinae) with some populations in Turkey. Journal of the Institute of Science and Technology. 11(2): 1489-1496.

Shivakumara, K.T., T. Venkatesan, M.C. Keerthi, P.R. Shashank, N. Pradeeksha, A. C. Polaiah, R. N. Reddy, P. L. Saran, & P. Manivel. 2021. Occurrence of *Pyrausta panopealis* on sweet basil *Ocimum basilicum* in India. Journal of Environmental Biology. 42: 265-270.

Wikström, B., P. Huemer, M. Mutanen, J. Tyllinen, & L. Kaila. 2020. *Pyralis cardinalis*, a charismatic new species related to *P. regalis* [Denis & Schiffermüller], 1775, first recognized in Finland (Lepidoptera: Pyralidae). Nota Lepidopterologica. 43: 337-364.

Xiang, L., K. Chen, & D. Zhang. 2021. Revision and phylogeny of the genus *Loxoneptera* Hampson, 1896 (Lepidoptera, Crambidae, Pyraustinae), based on morphology and molecular data. ZooKeys. 1036:75-98.

Scopariinae

Karisch, T. & L. Fowler. 2021. Description of the larva of *Helenoscoparia nigritalis* (Walker, 1855) (Pyraloidea: Crambidae: Scopariinae). Metamorphosis. 32:15-23.

Spilomelinae

Handfield, L. & D. Handfield. 2021. A revision of the Canadian species of the Genus *Herpetogramma* Lederer, 1863 (Lepidoptera: Crambidae: Spilomelinae: Herpetogrammatini), with descriptions of three new species. Bonn Zoological Bulletin. 70(1): 173-199. https://doi.org/10.20363/bzb-2021.70.1.173

Jeong, N., M. Kim, S.-S. Kim, S.-W. Choi, & I. Kim. 2021. Morphological, ecological, and molecular divergence of *Conogethes pinicolalis* from *C. punctiferalis* (Lepidoptera: Crambidae). Insects. 12(5): 455.

Ko, J.-H., T.-G. Lee, U. Bayarsaikhan, H.-U. Kim, B.-S. Park, & Y.-S. Bae. 2021. Review of genus *Palpita* (Lepidoptera: Crambidae: Spilomelinae) from Laos, part 1: A group. Journal of Asia-Pacific Biodiversity. https://doi. org/10.1016/j.japb.2021.01.002

Ko, J.-H., T.-G. Lee, U. Bayarsaikhan, Y.-B. Cha, B.-S. Park, & Y.-S. Bae. 2021. Review of genus *Palpita* Hübner (Lepidoptera: Crambidae: Spilomelinae) from Laos, part 2: B group. Journal of Asia-Pacific Biodiversity. https://doi. org/10.1016/j.japb.2021.07.007

Ko, J.-H., U. Bayarsaikhan, T.-G. Lee, Y.-B. Cha, J. K. Lee, & Y.-S. Bae. 2021. A new species of *Palpita* Hübner from Laos (Lepidoptera, Crambidae, Spilomelinae). Zootaxa. 4990(3): 577-582.

Lee, T.-G., B.-W. Lee, J.-H. Ko, Y.-B. Cha, H. Kim, B.-S. Park, & Y.-S. Bae. 2021. Two newly recorded species of Spilomelinae (Lepidoptera: Crambidae) from Korea. Journal of Asia-Pacific Biodiversity. 14(2): 271-274.

Matsui, Y., H. Naka, & U. Jinbo. 2021. DNA barcoding and morphology reveal a new cryptic species of *Nagiella* (Lepidoptera, Crambidae, Spilomelinae) from Japan. ZooKeys. 1023: 171-192.

Morse, D. H. 2021. Spider use of caterpillar shelters. Journal of Arachnology. 48: 284-287. [*Herpetogramma*]

Rosfiansyah, [no initial], S. Yagi, S. Tomura, & T. Hirowatari. 2021. A new species of the genus *Agrioglypta* Meyrick (Lepidoptera: Crambidae). Journal of Asia-Pacific Biodiversity. 14: 557-568. Seizmair, M. 2021. First records of the genus *Cirrhochrista* Lederer, 1883 (Lepidoptera, Crambidae, Spilomelinae) from the Arabian Peninsula with the description of *Cirrhochrista seminvea* [correct spelling *seminivea*] sp.n. Journal of Entomology and Zoology Studies. 9(2): 46-49.

Seizmair, M. 2021. Taxonomic and faunistic results on the Spilomelinae Guenée, 1854 (Lepidoptera: Crambidae) from the Southern Arabian Peninsula with descriptions of three new species. American Journal of Entomology. 5(1): 1-9.

Sohn, J.-C., S.-S. Kim, J.-M. Koo, & S.-W. Choi. 2021. Review of *Cotachena* Moore, 1885 (Lepidoptera: Crambidae, Spilomelinae) from Korea based on morphology and DNA barcodes. Journal of Asia-Pacific Entomology. 24(1): 383-389. https://doi.org/10.1016/j. aspen.2021.01.011

Vargas, H. A. 2021. A new fruit-boring species of *Rhectosemia* Lederer (Lepidoptera: Crambidae) feeding on *Solanum peruvianum* L. (Solanaceae) in the Atacama Desert. Journal of the Lepidopterists' Society. 75(3): 191-196.

Yang, Z., M. Ullah, J.-F. Landry, S. E. Miller, M. E. Rosati, & Y. Zhang. Reassessment of the moth genus *Bacotoma*, with a new species from Hainan Island. Insect Systematics & Evolution. 51: 384-407.

Geographical

Aarvik, L., B.Å. Bengt, H. Elven, P. Ivinskis, U. Jürivete, O. Karsholt, M. Mutanen, & N. Savenkov. 2021. Additions and corrections to the Nordic-Baltic Checklist of Lepidoptera. Norwegian Journal of Entomology. 68: 1-14.

Beshkov, S., C. W. Plant, A Nahirnić, A. King, & P. Jakšić. 2020. A contribution to knowledge of Balkan Lepidoptera: moths collected in May-June 2018 in Austria, Slovenia, Serbia, North Macedonia, and Albania. The Entomologists' Record and Journal of Variation. 132: 24-45.

Cock, M. J. W. & M. Kelly. 2020. Fortyfive new records of moths (Lepidoptera) from Tobago, W.I., increase the total species known to 400. Living World, Journal of Trinidad and Tobago Field Naturalists' Club. 2020: 43-58.

Dar, A.A. and K. Jamal. 2021. Moth (Insecta: Lepidoptera) fauna of Sariska Tiger Reserve, Rajasthan, India. Notulae Scientia Biologicae. 13(2): 10906.

Garre, M. J., J. Girdley, J. J. Guerrero, R. M. Rubio, & A. S. Ortiz. 2021. An annotated checklist of the Crambidae of the region of Murcia (Spain) with new records, distribution and biological data (Lepidoptera: Pyraloidea, Crambidae). Biodiversity Data Journal. 9: e69388. doi: 10.3897/BDJ.9. e69388

Gibson, L. D., C.V. Covell, Jr., & E. L. Laudermilk. 2021. Additions, deletions, and corrections to the Kentucky list of Lepidoptera. News of The Lepidopterists Society. 63(1): 24-32.

Gumhalter, D. & M. Kuĉinić. 2021. Contribution to the knowledge of the Croatian Pyraloidea fauna. Species reported from Biokovo Natural Park (Insecta: Lepidoptera). SHILAP Revista de Lepidoptera. 49 (193): 65-83.

Gumhalter, D., M. Kuĉinić, & Šašić. 2020. Data on pyraloid specimens (Lepidoptera: Pyraloidea) held in collections of CNHM in Zagreb, Croatia. Zootaxa. 4895(1): 37-66.

Huemer, P. 2020. Schmetterlinge (Lepidoptera) einer Submediterranen enclave südtirols (Fenner Schlucht, Magried), pp. 320-347. Wissenschaftliches jahrbuch der Tiroler Landemuseen. StudienVerlag, Innsbruck, Wien.

Krüger, M. 2020. Checklist of the Lepidoptera of southern Africa. Metamorphosis. 31(2): 1-201 Landry, B. Y. Basset, P. D. N. Hebert, & J.-M. Maes. 2020. On the Pyraloidea fauna of Nicaragua. Tropical Lepidoptera Research. 30(2): 93-102.

Nayak, A. 2021. A preliminary checklist of moths (Lepidoptera: Heterocera) from Gangajalghati, Bankura, West Bengal, India. Journal of Threatened Taxa. 13(9): 19310-19323.

Qi, M., H. Sun, X. Zuo, & H. Li. 2021. Annual report of new taxa for Chinese Lepidoptera in 2020. Biodiversity Science. 29(8): 1035-1039. [In Chinese with English Abstract]

Rao, B. S. K. & Sivaperuman, C. 2020. Annotated checklist of pyraloid moths (Lepidoptera: Pyraloidea) of Andaman and Nicobar Islands. Journal of the Andaman Science Association. 25(1): 15-26.

Sondhi, S., T. Karmakar, Y. Sondhi, & K. Kunte. 2021. Moths of Tale Wildlife Sanctuary, Arunachal Pradesh, India with seventeen additions to the moth fauna of India (Lepidoptera: Heterocera). Tropical Lepidoptera Research. 31: 1-53 (Suppl. 2).

Yepishin, V., Y. Khalaim, Y. Budashkin, O. Zhakov, V. Mushynskyi, & S. Novytskyi. 2021. New records of pyraloid moths (Lepidoptera: Pyraloidea) from different regions of Ukraine. Zootaxa. 5023(3): 366-388.

Biology

Cabral-de-Mello, D. C., M. Zrzavá, S. Kubíčkova, P. Rendón, & F. Marec. 2021. The role of satellite DNAs in genome architecture and sex chromosome evolution in Crambidae moths. Frontiers in Genetics. 12: 661417. doi: 10.3389/fgene.2021.661417

Nunes, J. 2021. New records of Lepidoptera hostplants from Portugal (Insecta: Lepidoptera). SHILAP Revista de Lepidopterologia. 49(193): 5-19.

Patil, A. H. & J. Sandesh. 2021. Noctur-

nal insect pollinator diversity and species richness in Ridge Gourd, *Luffa acutangular*. Applied and Natural Science Foundation. 13(3): 463-469. [features photos of spilomelines and some acentropines on flowers]

Singh, A. P. & G.C. Ramola. 2021. New host records of Lepidoptera defoliating Himalayan silver oak, *Quercus leucotrichophora* a. Camus and ring-cupped oak, *Q. glauca* Thunb. (Fagaceae) in Uttarakhand, Western Himalayas, India. Entomon. 46(1): 53-68.

Yaakop, S., A. David-Dass, U. S. Shaharuddin, S. Sabri, A. S. Badrulisham, & C. M. Z. Che-Radziah. 2020. Species richness of leaf roller and stem borers (Lepidoptera) associated with different paddy growth and first documentation of its DNA barcode. Tropical Agricultural Science. 43(4): 523-535.

Zhang, Z.-B., N.-N. Yin, J.-M. Long, Y.-K. Zhang, N.-Y. Liu, & J.-Y. Zhu. 2020. Transcriptome analysis of the pheromone glands in *Noorda blitealis* reveals a novel AOX group of the superfamily Pyraloidea. Journal of Asia-Pacific Entomology. 24 (2021): 110-119. https:// doi.org/10.1016/j.aspen.2020.11.007

Mitochondrial genomes

Guo, P. 2021. Next-generation sequencing yields the complete mitogenome of the stored nut moth, *Paralipsa gularis* Zeller (Lepidoptera: Pyralidae). Mitochondrial DNA Part B: Resources. 6(9): 2626-2627.

Jeong, N. R., M. J. Kim, J. S. Park, S. Y. Jeong, & I. Kim. 2021. Complete mitochondrial genomes of *Conogethes punctiferalis* and *C. pinicolalis* (Lepidoptera: Crambidae): genomic comparison and phylogenetic inference in Pyraloidea. Journal of Asia-Pacific Entomology. 24: 1179-1186.

Lu, Q., N. Zhou, & Z. Yang. 2021. Complete mitochondrial genome of *Ostrinia kasmirica* (Lepidoptera: Crambidae). Mitochondrial DNA Part B: Resources. 6(8): 2318-2318.

Liu, B., H. Sun, Q. Zhan, & Y. Gai. 2021. The complete mitochondrial genome of *Orthaga achatina* (Lepidoptera: Pyralidae). Mitochondrial DNA Part B: Resources. 6(3):821-823.

Pyraloid Enthusiasts

Please refer or forward the details to me about anyone who wishes to be put on the Pyraloid Planet distribution list.

If you have any suggestions, comments, and, more importantly, additions for next year's edition, please send to me ASAP, or during the year. The next deadline will be October/November 2022.

Name

David Agassiz Stacey Anderson J. E. F. Asselbergs Yang Seup Bae George J. Balogh Hans Bänziger Henry S Barlow Alejandro Barro Graziano Bassi Franziska Bauer Stella Beavan Vitor O. Becker Rebecca Bennik Will Bernstein Richard L. Brown Alain Cama Everett D. (Tim) Cashatt Danilo Cepeda Montero A.K.Chakravarthy Fugiang Chen José Clavijo Matthew Cock Alexandre-Pierre Cotty Srećko Ćurčić Willy De Prins Julian P. Donahue Xicui Du

email

david.agassiz@gmail.com Stacey.Anderson@aqis.gov.au JEF.Asselbergs@hetnet.nl baeys@incheon.ac.kr bugdr@att.net sangdao.banziger@cmu.ac.th hsbar@hotmail.com abarro@fbio.uh.cu graziano.bassi@alice.it franziska.bauer@senckenberg.de Stellabeavan@outlook.com vitor.o.becker@gmail.com mbennik@hotmail.com bugguycolorado@gmail.com moth@ra.msstate.edu alain.cama@orange.fr cashatt@museum.state.il.us museoent@uchile.cl chakravarthyakshay@gmail.com chenfg@ioz.ac.cn pepeclavijoa@gmail.com m.cock@cabi.org alexandre.cotty@gmail.com srecko.curcic@yahoo.com Willy.deprins@gmail.com julian.donahue@gmail.com lucy2073@sina.com

Marc Epstein Atousa Farahpour-Haghanih Guillermo Fernandez Clifford D. Ferris Vilhelmsen Flemming **Reinhard Gaedike Barry Goater** Kurt Grimm Chris Grinter Christian Guillermet **Bartholomew Hacobian** John Hawking James E. Hayden **Bob Heckford** John B. Heppner Alvaro Herrera Villalobos alherrer@inbio.ac.cr Robert J. B. Hoare Terence Hollingworth Martin Honey Marianne Horak Robin Howard Peter Huemer Utsugi Jinbo Ole Karsholt Akito Kawahara Gareth Edward King Nagaraj Kammar **Rajesh Kumar Gregor Kunert** Bernard Landry Jean-François Landry David Lees Théo Léger Guillaume Leraut Patrice Leraut Houhun Li Weichun Li Jiayu Liu Jean-Michel Maes Koen Maes **Richard Mally** Geoff Martin Edda Martinez **Hugh McGuinness** Mark Metz Eric Metzler Wolfram Mey Joël Minet Don Miller Scott Miller Andrew Mitchell Charlie Mitter Hideshi Naka Herb H. Neunzig Valentin Nidergas Matthias Nuss Eivind Palm Steven Passoa **Eugenie Phillips**

mepstein@cdfa.ca.gov

papiliona@gmail.com guillermo.fernandez@uv.es cdferris@uwyo.edu flemming.vilhelmsen@anarki.dk tinagma@msn.com lignaperda@myphone.coop kurtgrimm@bluewin.ch cgrinter@gmail.com chring@club-internet.fr barthacobian@optusnet.com.au J.Hawking@latrobe.edu.au jehayden63@gmail.com bob.heckford@btinternet.com jheppner@flmnh.ufl.edu HoareR@landcareresearch.co.nz alkibbir@gmail.com m.honey@nhm.ac.uk marianne.horak@csiro.au robin.lasdescargues@gmail.com p.huemer@tiroler-landesmuseen.at ujinbo@kahaku.go.jp okarsholt@snm.ku.dk kawahara@flmnh.ufl.edu sterrhinae@gmail.com rajsk0000@gmail.com rajesh.ento@gmail.com Gregor.Kunert@kbs-leipzig.de bernard.landry@ville-ge.ch Jean-francois.landry@canada.ca davil@nhm.ac.uk theo.leger@outlook.com guillaume.leraut@hec.edu pleraut@mnhn.fr lihouhun@nankai.edu.cn weichunlee@126.com fsliujiayu@163.com jmmaes@yahoo.com kvmaes@gmail.com spilomallynae@gmail.com G.Martin@nhm.ac.uk edda.martinez1@upr.edu hdmcguinness@gmail.com mark.metz@ars.usda.gov ehnmetzler@metzler.app wolfram.mey@mfn-berlin.de minet@mnhn.fr entdon@gmail.com MillerS@si.edu Andrew.Mitchell@austmus.gov.au cmitter@umd.edu chun@shushu.me hneunzig@bellsouth.net valentin.nidergas@gmail.com matthias.nuss@senckenberg.de Epalm32@gmail.com Steven.C.Passoa@aphis.usda.gov phillipsrodriguez@gmail.com

Colin W. Plant Jerry A. Powell Qi Mu-jie Rahul Ranjan S. Rathikannu Yingdang Ren Amanda Roe **Daniel Rubinoff** Michael Sabourin Akio Sasaki **Brian Scholtens Rob Schouten** Christian H. Schulze Andreas Segerer Jay Shaffer Stephanie Shank M. Shankara Murthy P. R. Shashank Ayuna A. Shodotova Thomas J. Simonsen Harsimran Singh (Dino) Frantisek Slamka M. Alma Solis Wolfgang Speidel Felix Sperling Hari Sutrisno Stephen Sutton Juergen Thiele Peter Ustjuzhanin Erik van Nieukerken **Héctor Vargas** Francesca Vegliante Flemming Vilhemsen David L. Wagner Terry Whitaker Steven Whitebread Richard A. Worth Chunsheng Wu Lijun Xu Zhaofu Yang Shen-Horn Yen Ping You James D. Young Dandan Zhang

cpauk1@ntlworld.com powellj@nature.berkeley.edu qimujie@163.com rranjan720@gmail.com rathikannu@gmail.com renyd@126.com amandaroe5@gmail.com rubinoff@hawaii.edu mothvet@yahoo.com scopar089@ybb.ne.jp scholtensb@cofc.edu rschouten@museon.nl christian.schulze@univie.ac.at Andreas.Segerer@zsm.mwn.de jshaffe1@gmu.edu sgshank53@gmail.com smurthyent@gmail.com spathour@gmail.com shodotova@mail.ru t.simonsen@nathist.dk dinotude.sym@gmail.com f.slamka@nextra.sk alma.solis@usda.gov speidel@zsm.mwn.de Felix.Sperling@ualberta.ca sutrisnohari@yahoo.com stephensutton7@gmail.com thithy@t-online.de petrust@mail.ru erik.vannieukerken@naturalis.nl havargas@uta.cl francesca.vegliante@senckenberg.de flemming.vilhelmsen@anarki.dk david.wagner@uconn.edu t.whitaker1@btinternet.com whitebread@one-name.org rworth@oda.state.or.us wucs@ioz.ac.cn zhilanmeng@qq.com yangzhaofu@nwsuaf.edu.cn shenhornyen@gmail.com vouping@snnu.edu.cn Jim.d.young@aphis.usda.gov zhdd61@163.com

